Development of Value Added Product from Chicory Roots

Dissertation 1 Report

Submitted by,

Dinesh Suryakant Patil

11714425 Programme – M.Tech. (Food Technology) Section – H1732

School of Agriculture

Lovely Professional University, Phagwara



Under the Guidance of Sawinder kaur

Coordinator

Department of Food

Technology and

Nutrition School of

Agriculture

Lovely Professional University, Phagwara

May 2017



CERTIFICATE

This is to certify that **Dinesh Suryakant Patil** (Registration No.11707850) has personally completed M.Tech. Dissertation 1 entitled, **"Development of Value Added Product from Chicory Roots"** under my guidance and supervision. To the best of my knowledge, the present work is the result of his original investigation and study. No part of Dissertation has ever been submitted for any other purpose at any University.

The project report is appropriate for the submission and the partial fulfilment of the conditions for the evaluation leading to the award of Master of Food Technology.

Date: 12-05 -2018

Signature of supervisor Sawinder Kaur Coordinator Department of Food Technology and Nutrition School of Agriculture Lovely Professional University Phagwara, Punjab, India

TABLE OF CONTENT

1. INTRODUCTION	
2. PROBLEMS BACKGROUND	6
3. REWIEW OF LITERATURE	
4. PROPOSED RESEARCH OBJECTIVE	
5. PROPOSED RESEARCH METHODOLOGY	
6. EXPECTED RESEARCH OUTCOME	
7. REFERENCES	

1. INTRODUCTION:

Chicory (Cichorium intybus L.) has several names it is also known as "Succory", "Hendibeh" or "Witloof" it's a wild edible plant consumed in Lebanon, Arab countries and in different parts of the World. The family Cichorium (Asteraceae) consists of 6 species with major distribution areas in Europe and Asia (Bais & Ravishankar 2001). In several Asteraceae, inulin, acts as a reserve carbohydrate in stems, tubers, and taproots, inulin is a β -2,1 linked fructose polymer with a terminal glucose residue (Arkel et al, 2012). Chicory is a woody perennial herb having around Im height with a fleshy taproot of up to 75 cm in length and large basal leaves (Gericke et al, 1997). From the past many years, chicory was grown by the ancient Egyptians used as a medicinal purpose, coffee substitute, and vegetable crop and was rarely used for animal feed. In the 1970s, it was discovered that the root of C. intybus contained inulin up to 40%, which has a minor impact on blood sugar and thus is suitable for Sugar patients (Judzentiene & badiene 2008) Up till now, C. intybus is developed for the production of inulin on an industrial scale (vergauwen et al, 2012). The name derived from Greek and Latin. Cichorium means field and intybus is Greek word means "to cut", because of the leaves, and from the Latin tubus to specify the hollow stem (European Medicines Agency 2013) Chicory can grow wild, in its natural environment, in fields, roadsides or home gardens. Chicory consists of the flower having blue color, roots and "wild" green leaves. Roots and Leaves are the edible parts of the plant. Chicory mainly eaten in cooked dishes mainly mixed with lemon juice, olive oil, salt and fried onions or raw in green salads (Ravishankar & Bais, 2001). Bushy edible plants are green leafy vegetables that are a excellent source of dietary fibers, minerals, vitamins, antioxidants and phytochemicals and contribute to the prevention of diseases such as cardiovascular problems, gastrointestinal problems, and diabetes (Allende et al., 2006; Barat et al., 2007). However, the utilization of wild edible plants has been declining lately due to ecological and economic factors and changes in lifestyle/cooking habits (Jeambey, 2005; Batal et al., 2009). Therefore, it is necessary to improve

the production and consumption of green leafy vegetables in local diets and find efficient preservation methods which are suitable for keeping these plants "fresh" with respect to good quality and nutritional content.

Chicory is a biyearly plant with several applications in the food industry. The roots of chicory are used in dried and roasted form and used for blending with coffee to add flavour; the fresh leaves are used in salads and vegetable dishes, and the chicory extracts are used for the production of invigo-rating beverages (Milala et al., 2010).

The wastes generated from the chicory (leaves and peel) are very hardly used in processing industry. The management of vegetable and fruit wastes seems to be advisable and is of growing scientific interest (Peschel et al., 2006). In recent studies it was demonstrated that the Cichorium species possesses both pro- and antioxidants (Papetti et al., 2006), and additional researchers have pointed to a phenolic fraction with potential antiradical activity of chicory (Gallori et al 2005). However, not only phenolic compounds can be responsible for the in free radical scavenging activity of chicory preparations. For example, it is proposed that sugars themselves, especially sucrose and fructans, can also act as radical scavengers in plant cells (Valluru et al., 2009). The previous data, in connection with a new scientific approach to radical scavenging activity as the major health-promoting property linked with dietary fibre (Goni et al., 1996), encouraged us to study non-inulin fraction of chicory by-products as potential health promoting agents improving physiological activity of inulin.

Cichorium intybus is a plant grown in Eurasia and in parts of Africa for its medicinal important, even with its long tradition of use; the plant is not described in the European Pharmacopoeia or in any official Pharmacopoeia of a European Union member state (European Medicines Agency, 2008). However, due to its common distribution, different parts of the plant have been used in conventional medicines globally (Akkola et al., 2012). Important phytochemicals are distributed throughout the plant, but the main contents are present in the root (Street et al., 2013). This study focuses on the economic and culturally important medicinal & Industrial uses of C. intybus.

2. PROBLEM BACKGROUND:

Root chicory has been grown in Europe as a coffee substitute, although chicory might have originated in France, Italy, and India. It studies shows that the root contains up to 20% inulin, a polysaccharide similar to starch. Chicory was commercialized but only with a limited use like Inulin extraction, Coffee Substitute. Mainly research's carried on chicory for the Inulin but it was found in studies that chicory has a many other chemical constituents like phenolic compounds present in chicory, antioxidant, antimicrobial activity. It can be used for treating various disorders such as diabetes and cardiovascular diseases. Therefore, the present research has been planned to development of value added products from chicory which can be commercialized. Development of products like Chicory juice, Chicory root powder from crude extract, fermented product as chicory root pickle, use of chicory flour in muffins as sugar and fat replacement.

3. REVIEW OF LITRATURE

3.1 Traditional uses of chicory:

Chicory has a long history of therapeutic use both in areas where it is indigenous and in areas where it has been introduced. Different preparations of this plant are in use to treat various symptoms and ailments. According to the European monograph, chicory roots having traditional uses includes relief from symptoms related to mild digestive disorders (abdominal fullness, flatulence, and slow digestion) and loss of appetite (European Medicines Agency 2012). During the digestive disorders and loss of appetite the use of a tea of the roots was already reported in 1938 by Madaus (European Medicines Agency, 2012.).

In purification of blood, catarrh of the stomach, jaundice the roots are used. The use of aqueous root extracts as a light-sensitive plant remedy for malaria was described in folkloric report prior to the wars in Afghanistan. The light-sensitive compounds of C. intybus root are sesquiterpene lactones lactucin and lactucopicrin antimalarial compounds have been identified (Bischoff et al. 2004). The roots are used as a bitter tonic or stomachic, mild laxative, to support liver function and diuresis (European Medicines Agency, 2012.). The flowers of the chicory plant are used as a herbal treatment of everyday ailments such as a tonic and appetite stimulant and as a treatment of, gastroenteritis, gallstones, sinus problems, bruises and cuts. (judzentiene 2008).

Chicory is traditionally used in slimming diets as a adjuvant. Traditionally used to promote renal elimination functions (Ministry of social affairs and solidarity 1990). Jigrine is a commercial product of India used for the treatment of various diseases of the liver and Chicory seeds are one of the main ingredient (Ahmed et al., 2003). Other parts of chicory are also used for liver disorders, namely, aerial parts in Bosnia and Herzegovina (Hanlidou et al., 2004) and roots in Serbia and India (Low et al., 2007).

3.1.1Traditional medicinal uses of chicory:

Country	Traditional use	Plant part	
Afghanistan	Malaria	Root	
Bosnia & Herzegovina	Diarrhea, Strengthening the prostate & other reproductive organs, pulmonary cancer, hangover, and purification of biliary tract.	Aerial part, flowers, roots	
Bulgaria	Cholagogue stimulant / gastric secretion, hypoglycemic	Roots, aerial part	
India	Liver disorders, Diabetes, Jaundice, liver enlargement, Rheumatism & gout, cough relief	Seed, Whole Plant, Root	
Italy	Blood Cleansing, High blood pressure, Blood purification, Italy Arteriosclerosis, Antiarthristis, Antipasmodic, Digestive		
Iran	Eupeptic, stomachic,	Whole plant	

Table no.1 Traditional medicinal uses of Chicory

	depurative, choleretic, laxative, hypotension, tonic, and antipyretic	
Jordan	Internal hemorrhage, sedative in typhoid	Whole plant
Pakistan	Diabetes	Root
South Africa	Jaundies, Tonic	Leaves, Steam , roots
Turkey	Cancer, Kidney stone, Wound healing, Hemorrhoids, urinary disorders	Root, Leaf, Aerial

Source: (Street et al, 2013)

3.2 Chemical Constituents:

It was clear that chicory leaves contain high level of protein (14.70%) ash (10.91) crude ether extract (3.68%) & crude fiber (16.78%) & had low concentration of total carbohydrates (68.50%) compared to with its roots. On the other hand, the roots of chicory were characterized by its high concentration of inulin (44.69%) (Monti et al.,2005). Crude protein range from 8.56 - 15.73% and 9.53 - 13.75% respectively.

in methanolic extracts Chicoric acid has been identified as the major compound of chicory (Table 2)

Compounds	Reference
Malic acid, Caffeic acid, 3-Caffeoylquinic	Carazzone et al., 2013
acid, 5-Caffeoylquinic acid, 4-Caffeoylquinic	
acid, cis-5-Caffeoylquinic acid, cis-Caftaric	
acid, trans-Caftaric acid, 5-Caffeoylshikimic	
acid, 5-p-Coumaroylquinic acid, Quercetin-3-	
O-glucuronide-7-O-(6"-Omalonyl)-glucoside,	
Kaempferol-3-O-glucosyl-7-O-(6"-Omalonyl)-	
glucoside, Dimethoxycinnamoyl shikimic acid,	
Kaempferol-3-O-sophoroside, Isorhamnetin-7-	
O-(6"-O-acetyl)-Glucoside, 5-O-	
Feruloylquinic acid, Dicaffeoyltartaric acid	
(chicoric acid), Kaempferol-7-O-glucosyl-3-O-	
(6"malonyl)-glucoside, Delphinidin-3-O-(6"-	
O-malonyl)-glucoside-5-O-glucoside,	
Cyanidin-3,5-di-O-(6"-O-malonyl)-Glucoside,	
3,5-Dicaffeoylquinic acid, Cyanidin-3-O-(6"-	
O-malonyl)-glucoside, Petunidin-3-O-(6"-O-	
malonyl)-Glucoside, Cyanidin, Cyanidin-3-O-	
galactoside, Cyanidin-3-O-(6"-O-acetyl)-	
glucoside, Malvidin-3-O-glucoside,	
Pelargonidin-3-O-monoglucuronide, 4-O-	
Feruloylquinic acid, Apigenin-7-O-glucoside,	
Chrysoeriol-3-O-glucoside, Tricin-3-O-	
glucoside, 1,3-Dicaffeoylquinic acid, 1,4-	
Dicaffeoylquinic acid, 3,4-Dicaffeoylquinic	
acid, Quercetin-7-O-galactoside, Quercetin-3-	
O-(6"-O-malonyl)-Glucoside, Quercetin-7-O-	

(6"-O-acetyl)-glucoside, Quercetin-7-O-	
glucuronide, Quercetin-7-O-(6"-O-acetyl)-	
glucoside, Kaempferide glucuronide,	
Kaempferol-7-O-glucoside, Kaempferol-7-O-	
rutinoside, Quercetin-7-O-p-	
coumaroylglucoside, Isorhamnetin-7-O-	
neohesperidoside, Kaempferol-7-O-(6"-O-	
malonyl)-glucoside, Kaempferol-7-O-	
glucuronide, Kaempferide-3-O-(6"-O-	
malonyl)-Glucoside, Kaempferol-3-O-	
glucuronide, Kaempferol-3-O-glucuronide-7-	
Oglucoside, Kaempferol-3-O-(6"-O-malonyl)-	
Glucoside, Kaempferol-3-O-glucoside,	
Myricetin-7-O-(6"-O-malonyl)-glucoside,	
Kaempferol-7-O-neohesperidoside,	
Kaempferol-7-O-(6"-O-acetyl)-glucoside,	
Kaempferol-3-O-(6"-O-acetyl)-glucoside.	
Lactucin, Lactucopicrin, 8-Deoxylactucin,	Kisiel and K. Zieli´nska 2001
Jacquilenin, 11β ,13-Dihydrolactucin,	
Crepidiaside B, Cyanidin 3-O-p-(6-O-	
malonyl)-Dglucopyranoside, $3,4\beta$ -Dihydro-15-	
dehydrolactucopicrin, Magnolialide,	
Ixerisoside D, Loliolide, Cichorioside B,	
Sonchuside A, Artesin, Cichoriolide,	
Sonchuside C, Cichopumilide	
Delphinidin 3,5-di-O-(6-O-malonyl-β-D-	Norbak et al., 2002
glucoside), Delphinidin 3-O-(6-O-malonyl- β -	
Dglucoside)-5-O- β -D-glucoside, Delphinidin	
3-O- β -D-glucoside-5-O-(6-O-malonyl- β -D-	
L	1

glucoside), Delphinidin 3,5-di-O- β -D-	
glucoside, 3-O-p-Coumaroyl quinic acid.	
Oxalic acid, Shikimic acid, Quinic acid,	Papetti et al., 2013
Succinic acid	
	Bridle et al., 1984
Cyanidin 3-O- β -(6-O-malonyl)-	
Dglucopyranoside, Cyanidin 3-O-p-(6-O-	
malonyl)-Dglucopyranoside	
(7S,8R)-32-Demethyl	Malarz et al., 2013
dehydrodiconiferylalcohol-3⊡-O-β-	
glucopyranoside, 4,5-Dicaffeoylquinic acid,	
Crepidiaside A,	
Cichoralexin	Monde et al., 1990
Putrescine, Spermidine, β -Sitosterol,	Krebsky et al., 1999
Campesterol, Stigmasterol	

The major fractions are comprised by aliphatic compounds and their derivatives and terpenoids comprise minor constituents of the plant. The flowers of chicory contain saccharides, methoxycoumarin cichorine, essential oils, flavonoids (Judzentiene & Badiene., 2008) and the blue color of the perianth contributing anthocyanins (Wang & Cui 2011). Octane, hexadecane, pentadecanone, n-nonadecane and a tentatively identified compound have been found as principal volatile components (Badiene et al., 2008)

The mineral content of chicory root was found to be relatively less than chicory levaes. The macro elements present in chicory are Ca, K, Na, Mg and the micro elements Fe, Zn, Mn, Cu (Schittenhelm 2001)

3.3 Phenolic compound content as methanolic extract:

Yield of methanolic extract were about 23.16% in leaves & 10.75% in roots. Leaves : (Mona et al. 2004)

Methanolic Extract Yield : 23.16%

Total Phenolic Content : 26.4% (+/- 1.05)

Phenolic compound : Gallic Acid (1.96%), Protocatechuic Acid (2.50%), Chlorogenic Acid (17.84%), P. hydroxybenzoic Acid (11.04%), Caffeic Acid (35.22%), Iso Vanillic Acid (1.97%),
P. Coumaric Acid (9.65%), Other Compounds / Unknown compounds (19.46%)

Roots : (Mulinacci et al. , 2001)

Methanolic Extract Yield : 10.75%

Total Phenolic Content : 20.0% (+/- 0.90)

Phenolic compound: Protocatechuic Acid (1.77%), Chlorogenic Acid (10.85%), Caffeic Acid (24.36%), m.coumaric acid (27.90%), p.cumaric acid (25.03%),Unknown Compounds (10.09%)

3.4 Pharmacological activity:

3.4.1Antimicrobial activity:

The antibacterial activity of the organic acid-rich extract of fresh red chicory (C. intybus var. sylvestre) was tested against periodontopathic bacteria including Streptococcus mutans, Actinomyces naeslundii, and Prevotella intermedia. The compounds identified from the active extract include quinic acid, succinic acid, oxalic acid and shikimic acid. All of the organic acids were found to decrease biofilm formation and adhesion of bacteria to the cells, with different levels of efficacy. These compounds also induced biofilm disruption and detachment of dead cells for the cultured substratum (Papetti et al, 2000).

3.4.2 Antimalarial Activity:

The light-sensitive sesquiterpene lactones Lactucin and Lactucopicrin, extracted from Cichorium intybus roots, as antimalarial compounds. Both substances exhibited in vitro, activity against a HB3 clone of a strain Honduras-1 of Plasmodium falciparum. Inhibitory lactone concentrations that completely prevented parasite growth in cell cultures were 10 μ g/ml for Lactucin and 50 μ g/ml for Lactucopicrin (Bischoff et al, 2004).

3.4.3 Antioxidant Activity:

The DPPH radical scavenging activity of a polyphenols-rich fraction of C. intybus has been investigated (Heimler et al., 2009). The anti- and prooxidant activities of Cichorium species were studied in chemical as well as biological systems. In the case of chemical systems, the antioxidant activity of water-soluble compounds in C. intybus var. silvestre was established in the coupled model of linoleic acid and β -carotene. A pro-oxidant activity of some of the chemical components was recorded initially which notably diminished with time and/or thermal treatment. Thereafter, the antioxidant activity of the raw juice and its fractions persisted. The molecular weight ranges of the antioxidant fractions of raw juice were also identified based on dialysis (Papetti et al., 2006).

Two varieties of chicory, namely, C. intybus var. Silvestre and C. intybus var. foliosum, have been investigated for their antioxidant (antiradical) activities in two distinct biological systems. The lipi peroxidation assay has been carried out on microsomemembranes of rat hepatocytes after the induction of oxidative damage by carbon tetrachloride. The antiradical activity was expressed as the protective activity against lipid peroxidation and calculated as the percentage decrease in hydroperoxide degradation products. The second biological system used was the cultures of S. aureus after treatment with cumene hydroperoxide. The percentage increase of growth of bacteria was noted after the treatment with juices of chicory varieties. In both systems, the juices of chicory varieties showed strong antiradical activities (Gazzani et al., 2000). Red chicory (C. intybus var. silvestre) was studied for its polyphenol content and the antioxidant activity was evaluated by using the synthetic 2,2-diphenyl-1-(2,4,6-trinitrophenyl) hydrazyl radical and three model reactions catalyzed by pertinent enzymatic sources of reactive oxygen species, namely, xanthine oxidase, myeloperoxidase, and diaphorase. Total phenolics were significantly correlated with the antioxidant activity evaluated with both the synthetic radical and the enzyme-catalyzed reactions. Onamolar basis, redchicory phenolics were as efficient as Trolox (reference compound) in scavenging the synthetic radical (Lavelli 2008). The aqueous-alcoholic extracts of the aerial parts of C. intybus also inhibited xanthine oxidase enzyme dose dependently (Pieroni et al., 2002). In another study, alongwithDPPHradical scavenging activity, C. intybus also exhibited inhibition of hydrogen peroxide and chelation of ferrous ion (El & Karakaya, 2004).

3.4.4 Antidiabetic effects:

An ethanolic extract of Cichorium intybus herb for its anti-diabetic activity on male Sprague Dawley rats treated streptozotocin. A dose of 125 mg/kg body weight influenced oral glucose tolerance test and the same amount given orally for 14 days reduced serum glucose by 20%, triglycerides by 91% and cholesterol by 16%. No changes in the insulin secretion were observed during the experiment, whereas hepatic Glc-6-Pase activity was markedly reduced (Pushparaj et al., 2007).

3.4.5 Anti-Inflammatory Activity :

The inhibition of TNF- α mediated cyclooxygenase (COX) induction by chicory root extracts was investigated in the human colon carcinoma (HT 29) cell line.Theethyl acetate extract inhibited the production of prostaglandin E2 (PGE2) in a dose-dependent manner. TNF- α mediated induction of COX-2 expression was also suppressed by the chicory extract (Cavin et al., 2005).

3.4.6 Antitumor activity:

A tumour-inhibitory effect of an ethanolic extract of chicory root against Ehrlich ascites carcinoma in mice. Significant results were obtained at doses from 300 to 700 mg/kg (Hazra et al., 2002).

3.4.7 Hepatoprotective Activity:

The folkloric use of C. intybus as a hepatoprotectant has been well documented. It is one of the herbal components of Liv-52, a traditional Indian tonic used widely for hepatoprotection. In a randomized, doubleblind clinical trial conducted on cirrhotic patients, Liv-52 medication reduced the serum levels of hepatic enzymes, namely, alanine aminotransferase and aspartate aminotransferase. It also reduced the Child-Pugh scores and ascites significantly (Fallah et al., 2005). Another polyherbal formulation, Jigrine, contains the leaves of C. intybus as one of its 14 constituents. Jigrine was evaluated for its hepatoprotective activity against galactosamine-induced hepatopathy in rats. The pretreatment of male Wistar-albino rats with jigrine significantly reduced the levels of aspartate transaminase, alanine transaminase, and urea and increased the levels of blood and tissue glutathione. Histopathological examination of the liver revealed that jigrine pretreatment prevented galactosamine toxicity and caused a marked decrease in inflamed cells (Aqil et al., 2005).

The aqueous-methanolic extract of the seeds of C. intybus has been investigated for the hepatoprotective activity against acetaminophen and carbon tetrachloride-induced liver damage inmice. It was found to decrease both the death rate and the serum levels of alkaline phosphatase, glutamyl oxaloacetate transaminase, and glutamyl pyruvate transaminase (Gilani & janbaz

1994). In analogous studies, the antihepatotoxic activity of the alcoholic extract of the seeds and aqueous extracts of the roots and root callus of C. intybus was estimated. The oral administration of these extracts in albino rats led to amarked decrease in the levels of hepatic enzymes. Also, histopathological examination of the liver showed no fat accumulation or necrosis after the treatment (Ahmed et al., 2003). Similar studies have established the hepatoprotective effect of esculetin, a phenolic compound, and cichotyboside, a guaianolide sesquiterpene glycoside reported from C. intybus (Khan et al., 2008).

The phenolic acid-rich seed extract of C. intybus was evaluated for its efficacy against hepatic steatosis in vitro and in vivo. The in vitro model of hepatic steatosis was created by incubation of the HepG2 cells with oleic acid leading to intracellular accumulation of fat. The seed extract was effective in decreasing the deposited fat from the cells in case of administration after the initial fat deposition (i.e., non simultaneous administration with oleic acid). However, in case of simultaneous administration of seed extract and oleic acid, the extract could not protect the cells from steatosis except at very high doses.

The extract also led to the increased release of glycerol (an indicator of triglyceride degradation) in steatotic cells. In case of nonsimultaneous administration, the extract was found to upregulate the expression of SREBP- 1c and PPAR- α genes leading to restoration of normal levels of corresponding proteins. In the in vivo model of hepatic steatosis, namely, diabetic rats, treatment with seed extract resulted in significant decrease in fat accumulation and fibrosis (Khaghania et al., 2013). The hepatoprotective activity of C. intybus has been correlated to its ability to inhibit the free radical mediated damage. A fraction prepared from the ethanolic extract of the leaves was assessed for preventive action on the free radical mediated damage to the deoxyribose sugar of the DNA (obtained from calf thymus). A dose-dependent decrease in the DNA damage was observed in the present assay (Sultana et al., 1995).

3.5 Food technological application:

Applications of Powder chicory root in some foods.

- 1) Beverage: Coffee
- 2) Crackers
- 3) Frozen Desserts
- 4) Bakery Food: Bread
 - 1) Beverages (Coffee) :

The powder of roasted chicory was used to replace 50-100% coffee.

Chicory is used as substitute for coffee.

Also reported that Nescafe blend chicory root(50%) had coarse perforated and light colour granules. (Mona et al., 2009)

2) Crackers :

The cracker were prepared according to the recipe and method (Adb-El-Rahim et al., 2003)

Chicory used as a Wheat flour (10%) and Fat replacer (25%).

3) Frozen Desserts: Yoghurt –Ice Cream

In yoghurt ice cream preparation the different trials taken for the improving the Colour, Flavour, Texture, Sweetness, Hardness, Melting rate etc.

The level of chicory root extract increases the score of colour of product was increased.

Chicory root extract shows positive effect on the flavour . Increase in the level of dried chicory root extract, the flavour of the product also increased.

Increase in the level of dried chicory root extract the texture increases.

In case of sweetness dried chicory root extract increases the score of sweetness.

Positive effect observed in case of increasing the hardness of yoghurt ice-cream.

Increase in chicory root extract decrease in melting rate.

Over all acceptability results shows that the chicory affects positively on all factors. (Kumar et al., 2017)

4) Bakery products : Rey Bread

In rey bread the chicory powder is used 9% for the improvement of fructans content.

The use of chicory powder is also affect on increasing consumer acceptance by its intrinsic aroma and sweet aftertaste.

It is necessary to indicate, that fructans concentration in bread could be affected not only by pH or acidity, but of increased processing temperature too, as higher than 90°C. (Gudreniece et al., 2017)

3.6. Removing the Bitterness

The bitter substances are primarily the two sesquiterpene lactones lactucin and lactucopicrin. Other ingredients are aesculetin, aesculin, cichoriin, umbelliferone, scopoletin, 6,7dihydrocoumarin, and further sesquiterpene lactones and their glycosides.(Bais et al., 2001) There are two different methods of debittering of chicory

- 1) Soaking
- 2) Roasting

Table :3 Effect of different treatment for removal of bitterness of chicory

Treatments	Time of		Bitter taste	Loss of	
	Soaking And	Inulin%		inulin%	
	roasting				
Control	-	44.69±0.27	+++++	-	
Soaking in	24hrs.	35.99±0.40	Slightly	19.47	
water at a ratio					
of 1:4 at R.T.					
Soaking in	1hrs.	37.61±0.54	+++	15.84	
water at a ratio	2hrs.	29.89±0.28	++	33.12	
1:4 at temp.	3hrs.	24.19±0.38	+	45.87	
45°C					
Soaking in	24hrs.	32.34±0.80	++	27.63	
Citric acid	48hrs.	29.55±0.42	+	33.87	
solution 0.75%	72hrs.	15.02 ± 0.28	-	66.39	
1:10					
Soaking in	2hrs.	43.57±0.42	+++++	2.51	
Ethanol 80%	3hrs.	42.82±0.29	+++++	4.18	
1:10	24hrs.	38.90±0.12	+++	12.96	
Roasting at	20min	38.05±0.08	++	14.86	
140°C	30min	37.51±0.38	+	16.07	
Roasting	30min	39.13±0.37	+	12.44	

at140°C + 10%		
treacle & 10%		
oil		

Source: Mona et al., 2009

The processs of debittering of chicory causes a loss in Inulin concentration, this reduction was based on type of method used for debittering. There are the different types of debittering are shown (Table 3).

As table shows that the reduction in percentage of inulin was considerably higher in soaking treatment than roasting. This could be due to the solubility of inulin in water and the low pH value of citric acid and its effect on inulinase enzyme activity.

4. PROPOSED RESEARCH OBJECTIVE

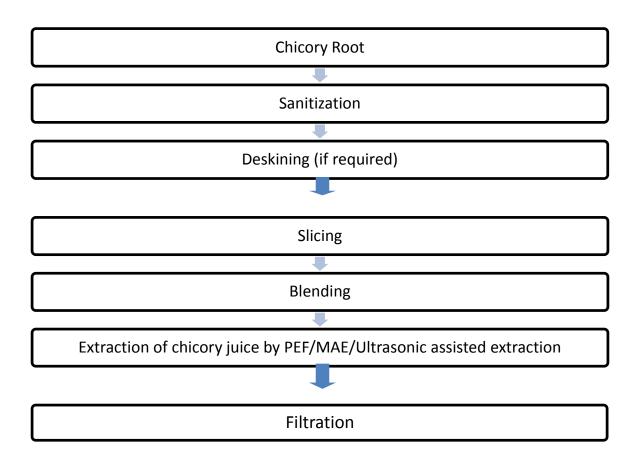
- 1) Preparation of Chicory Root Juice
- 2) Preparation of Chicory Root Powder by using crude juice extract
- 3) Preparation of Chicory Pickle as a source of Dietary fiber
- 4) Preparation of sugar and fat reduced muffins using Chicory Flour

5.0 PROPOSED RESEARCH METHODOLOGY:

5.1 Detailed plan work

5.1.1 Objective 1: Preparation / Extraction of chicory root juice

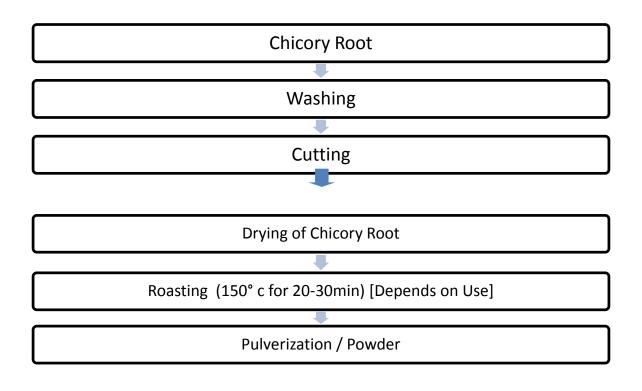
In contrast to the conventional inulin extraction process, which requires high temperature and long time, alternative technologies have been carried out, including supercritical carbon dioxide (CO2) ultrasound, simultaneous ultrasonic/microwave and pulsed electric field (PEF) assisted extraction (Vorobiev et al., 2013)



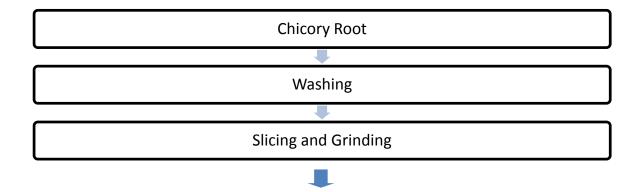
Proximate analysis:

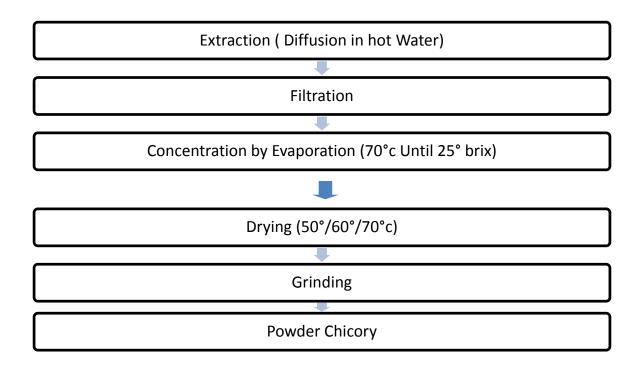
- Dry mass
- Soluble matter
- Inulin
- Carbohydrate
- Pectin
- Protein
- pH
- Viscosity
- Jucie Purity
- Antioxidant Activity

5.1.2 Objective 2: Preparation of Chicory Root Powder:



5.2.2.

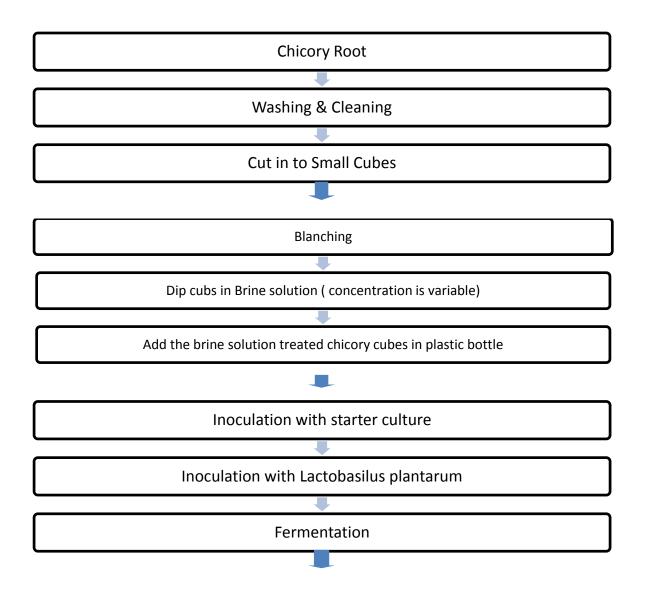


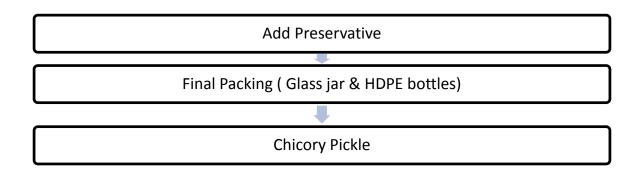


Proximate analysis:

- Concentration of pulp
- Loss of Weight during Drying
- Equilibrium moisture content
- Water Activity
- Sugar Concentration
- % of Inulin before & after drying
- Glucose and Fructose
- Moisture determination
- Microbial analysis
- Density
- Sensory Analysis

5.1.3 Objective 3: Preparation of Chicory Pickle as a source of Dietary fiber:





Proximate analysis:

(Taken as a reference for processing and testing -Mohapatra et al.,2017)

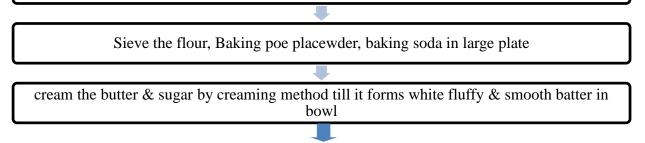
- Lactic acid production
- Starch
- Total Sugar
- Crude protein
- Fat
- Shelf Life
- Storage Condition Require
- Sensory (Bitterness)

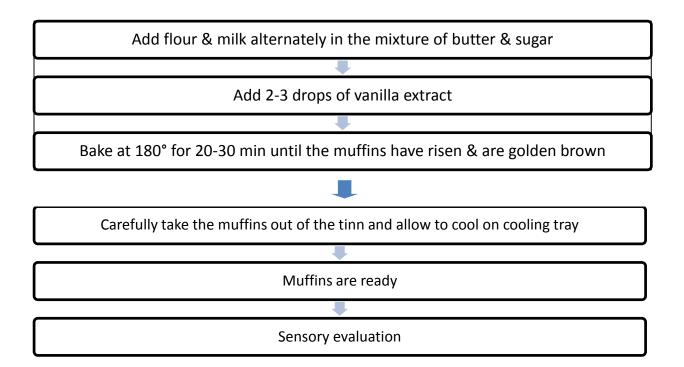
5.1.4 Objective 4: Preparation of sugar and fat reduced muffins using Chicory Flour:

A control recipe was prepared according to the literature (Hui & Corke, 2006) and slightly modified (Table), where sugar and fat was replaced by Chicory Flour.

Wheat	Sugar	Baking	Salt	Skim	Oil	Liquid	Water
Flour		Powder		milk		whole	
				powder		egg	
115.3	69.2	5.8	1.4	8.7	57.6	34.6	57.6
115.3	65.2	5.8	1.4	8.7	53	34.6	57.6
115.3	60.2	5.8	1.4	8.7	48	34.6	57.6
115.3	55.2	5.8	1.4	8.7	45	34.6	57.6
115.3	51.2	5.8	1.4	8.7	43	34.6	57.6
115.3	46.2	5.8	1.4	8.7	39	34.6	57.6

Take White flour (gluten free) and Chicory floiur (different % ratio)





Proximate analysis:

- A) Sensory Properties (9 Point Hedonic Scale)
- **B)** Physical Analysis
- Diameter
- Thickness
- Density
- Texture profile Analysis
- Nutritional Analysis :
- Ash Content (Ranganna, 2012)
- Moisture Content (Ranganna, 2012)
- Fat Content (Ranganna, 2012)
- Protein Content (Ranganna, 2012)
- Carbohydrates
- Total Starch analysis (AACC,1995).
- 1) Calcium
- 2) Phosphorus

3) Iron

6. EXPECTED RESEARCH OUTCOME

1. Developing the products from chicory root which is reach in Inulin as well rich in other phenolic compounds present in chicory

2. Developing the products which can be commercialized on industrial scale.

3. Developing the Fermented product as a source of dietary fiber.

4. Developing the product from chicory which have high nutritional value as well as having good taste, flavor, color, and overall acceptability.

7. REFERENCES:

- Bais HP, Ravishankar GA. Cichorium intybus L-cultivation, processing, utility, value addition and biotechnology, with an emphasis on current status and future prospects. Journal of the Science of Food and Agriculture. 2001 Apr 1;81(5):467-84.
- van Arkel J, Vergauwen R, Sévenier R, Hakkert JC, van Laere A, Bouwmeester HJ, Koops AJ, van der Meer IM. Sink filling, inulin metabolizing enzymes and carbohydrate status in field grown chicory (Cichorium intybus L.). Journal of plant physiology. 2012 Oct 15;169(15):1520-9.
- 3) Van Wyk BE, Oudtshoorn BV, Gericke N. Medicinal plants of South Africa. Briza; 1997.
- Judžentienė A, Būdienė J. Volatile constituents from aerial parts and roots of Cichorium intybus L.(chicory) grown in Lithuania. chemija. 2008 Jun 1;19(2).
- Street RA, Sidana J, Prinsloo G. Cichorium intybus: traditional uses, phytochemistry, pharmacology, and toxicology. Evidence-Based Complementary and Alternative Medicine. 2013 Nov 26;2013.
- Allende A, Tomas-Barberan FA, Gil MI. Minimal processing for healthy traditional foods. Trends in Food Science & Technology. 2006 Sep 30;17(9):513-9.
- Rico D, Martin-Diana AB, Barat JM, Barry-Ryan C. Extending and measuring the quality of fresh-cut fruit and vegetables: a review. Trends in Food Science & Technology. 2007 Jul 31;18(7):373-86.
- Jeambey Z, Johns T, Talhouk S, Batal M. Perceived health and medicinal properties of six species of wild edible plants in north-east Lebanon. Public health nutrition. 2009 Oct;12(10):1902-11.
- Jurgonski A, Milala J, Juskiewicz J, Zdunczyk Z, Król B. Composition of chicory root, peel, seed and leaf ethanol extracts and biological properties of their non-inulin fractions. Food Technology and Biotechnology. 2011 Jan 1;49(1):40.
- 10) Peschel W, Sánchez-Rabaneda F, Diekmann W, Plescher A, Gartzía I, Jiménez D, Lamuela-Raventos R, Buxaderas S, Codina C. An industrial approach in the search of natural antioxidants from vegetable and fruit wastes. Food Chemistry. 2006 Jul 31;97(1):137-50.

- 11) Papetti A, Daglia M, Grisoli P, Dacarro C, Gregotti C, Gazzani G. Anti-and pro-oxidant activity of Cichorium genus vegetables and effect of thermal treatment in biological systems. Food chemistry. 2006 Jul 31;97(1):157-65.
- 12) Innocenti M, Gallori S, Giaccherini C, Ieri F, Vincieri FF, Mulinacci N. Evaluation of the phenolic content in the aerial parts of different varieties of Cichorium intybus L. Journal of Agricultural and Food Chemistry. 2005 Aug 10;53(16):6497-502.
- 13) Van den Ende W, Valluru R. Sucrose, sucrosyl oligosaccharides, and oxidative stress: scavenging and salvaging?. Journal of Experimental Botany. 2008 Nov 26;60(1):9-18.
- 14) Larrauri JA, Goni I, Martín-Carrón N, Rupérez P, Saura-Calixto F. Measurement of healthpromoting properties in fruit dietary fibres: antioxidant capacity, fermentability and glucose retardation index. Journal of the Science of Food and Agriculture. 1996 Aug 1;71(4):515-9.
- 15) European Medicines Agency, "Assessment report on Cichorium intybus L., radix," EMA/HMPC/113041/2010, 2013.
- 16) Süntar I, Akkol EK, Keles H, Yesilada E, Sarker SD, Baykal T. Comparative evaluation of traditional prescriptions from Cichorium intybus L. for wound healing: stepwise isolation of an active component by in vivo bioassay and its mode of activity. Journal of ethnopharmacology. 2012 Aug 30;143(1):299-309
- 17) Bischoff TA, Kelley CJ, Karchesy Y, Laurantos M, Nguyen-Dinh P, Arefi AG. Antimalarial activity of Lactucin and Lactucopicrin: sesquiterpene lactones isolated from Cichorium intybus L. Journal of ethnopharmacology. 2004 Dec 31;95(2):455-7.
- 18) "Anonymous 2012", European Medicines Agency, 2012; 1:14.
- 19) Hanlidou E, Karousou R, Kleftoyanni V, Kokkini S. The herbal market of Thessaloniki (N Greece) and its relation to the ethnobotanical tradition. Journal of Ethnopharmacology. 2004 Apr 30;91(2):281-99.
- 20) Ahmed B, Al-Howiriny TA, Siddiqui AB. Antihepatotoxic activity of seeds of Cichorium intybus. Journal of ethnopharmacology. 2003 Aug 31;87(2):237-40.
- 21) Pushparaj PN, Low HK, Manikandan J, Tan BK, Tan CH. Anti-diabetic effects of Cichorium intybus in streptozotocin-induced diabetic rats. Journal of Ethnopharmacology. 2007 May 4;111(2):430-4.

- 22) Street RA, Sidana J, Prinsloo G. Cichorium intybus: traditional uses, phytochemistry, pharmacology, and toxicology. Evidence-Based Complementary and Alternative Medicine. 2013 Nov 26;2013.
- 23) Monti A, Amaducci MT, Pritoni G, Venturi G. Growth, fructan yield, and quality of chicory (Cichorium intybus L.) as related to photosynthetic capacity, harvest time, and water regime. Journal of Experimental Botany. 2005 Apr 4;56(415):1389-95.
- 24) Carazzone C, Mascherpa D, Gazzani G, Papetti A. Identification of phenolic constituents in red chicory salads (Cichorium intybus) by high-performance liquid chromatography with diode array detection and electrospray ionisation tandem mass spectrometry. Food Chemistry. 2013 Jun 1;138(2):1062-71.
- 25) Kisiel W, Zielińska K. Guaianolides from Cichorium intybus and structure revision of Cichorium sesquiterpene lactones. Phytochemistry. 2001 Jun 30;57(4):523-7.
- 26) Nørbæk R, Nielsen K, Kondo T. Anthocyanins from flowers of Cichorium intybus. Phytochemistry. 2002 Jun 30;60(4):357-9.
- 27) Papetti A, Mascherpa D, Carazzone C, Stauder M, Spratt DA, Wilson M, Pratten J, Ciric L, Lingström P, Zaura E, Weiss E. Identification of organic acids in Cichorium intybus inhibiting virulence-related properties of oral pathogenic bacteria. Food chemistry. 2013 Jun 1;138(2):1706-12.
- 28) Bridle P, Loeffler RT, Timberlake CF, Self R. Cyanidin 3-malonylglucoside in Cichorium intybus. Phytochemistry. 1984 Jan 1;23(12):2968-9.
- 29) Malarz J, Stojakowska A, Szneler E, Kisiel W. A new neolignan glucoside from hairy roots of Cichorium intybus. Phytochemistry Letters. 2013 Feb 28;6(1):59-61.
- 30) Monde K, Oya T, Takasugi M, Shirata A. A guaianolide phytoalexin, cichoralexin, from Cichorium intybus. Phytochemistry. 1990 Jan 1;29(11):3449-51.
- 31) Krebsky EO, Geuns JM, De Proft M. Polyamines and sterols in Cichorium heads. Phytochemistry. 1999 Feb 24;50(4):549-53.
- 32) Schittenhelm S. Effect of sowing date on the performance of root chicory. European journal of agronomy. 2001 Nov 30;15(3):209-20.
- 33) Mona IM, Wafaa AA, Elgindy AA. Chemical and Technological Studies on Chicory (Cichorium Intybus L) and Its Applications in Some Functional Food.

- 34) Mulinacci N, Innocenti M, Gallori S, Romani A, La Marca G, Vincieri FF. Optimization of the chromatographic determination of polyphenols in the aerial parts of Cichorium intybus L. Chromatographia. 2001 Oct 1;54(7):455-61.
- 35) Gazzani G, Daglia M, Papetti A, Gregotti C. In vitro and ex vivo anti-and prooxidant components of Cichorium intybus. Journal of pharmaceutical and biomedical analysis. 2000 Aug 1;23(1):127-33.
- 36) Heimler D, Isolani L, Vignolini P, Romani A. Polyphenol content and antiradical activity of Cichorium intybus L. from biodynamic and conventional farming. Food Chemistry. 2009 Jun 1;114(3):765-70.
- 37) Lavelli V. Antioxidant activity of minimally processed red chicory (Cichorium intybus L.) evaluated in xanthine oxidase-, myeloperoxidase-, and diaphorase-catalyzed reactions. Journal of agricultural and food chemistry. 2008 Aug 5;56(16):7194-200.
- 38) Pieroni A, Janiak V, Dürr CM, Lüdeke S, Trachsel E, Heinrich M. In vitro antioxidant activity of non-cultivated vegetables of ethnic Albanians in southern Italy. Phytotherapy Research. 2002 Aug 1;16(5):467-73.
- 39) Nehir El S, Karakaya S. Radical scavenging and iron-chelating activities of some greens used as traditional dishes in Mediterranean diet. International Journal of Food Sciences and Nutrition. 2004 Feb 1;55(1):67-74.
- 40) Pushparaj PN, Low HK, Manikandan J, Tan BK, Tan CH. Anti-diabetic effects of Cichorium intybus in streptozotocin-induced diabetic rats. Journal of Ethnopharmacology. 2007 May 4;111(2):430-4.
- 41) Cavin C, Delannoy M, Malnoe A, Debefve E, Touche A, Courtois D, Schilter B. Inhibition of the expression and activity of cyclooxygenase-2 by chicory extract. Biochemical and Biophysical Research Communications. 2005 Feb 18;327(3):742-9.
- 42) Hazra B, Sarkar R, Bhattacharyya S, Roy P. Tumour inhibitory activity of chicory root extract against Ehrlich ascites carcinoma in mice. Fitoterapia. 2002 Dec 31;73(7):730-3.
- 43) Huseini HF, Alavian SM, Heshmat R, Heydari MR, Abolmaali K. The efficacy of Liv-52 on liver cirrhotic patients: a randomized, double-blind, placebo-controlled first approach. Phytomedicine. 2005 Sep 15;12(9):619-24.

- 44) Najmi AK, Pillai KK, Pal SN, Aqil M. Free radical scavenging and hepatoprotective activity of jigrine against galactosamine induced hepatopathy in rats. Journal of ethnopharmacology. 2005 Mar 21;97(3):521-5.
- 45) Gilani AH, Janbaz KH. Evaluation of the liver protective potential of Cichorium intybus seed extract on acetaminophen and CCl4-induced damage. Phytomedicine. 1994 Dec 1;1(3):193-7.
- 46) Ahmed B, Khan S, Masood MH, Siddique AH. Anti-hepatotoxic activity of cichotyboside, a sesquiterpene glycoside from the seeds of Cichorium intybus. Journal of Asian natural products research. 2008 Mar 1;10(3):218-23.
- 47) Ziamajidi N, Khaghani S, Hassanzadeh G, Vardasbi S, Ahmadian S, Nowrouzi A, Ghaffari SM, Abdirad A. Amelioration by chicory seed extract of diabetes-and oleic acid-induced nonalcoholic fatty liver disease (NAFLD)/non-alcoholic steatohepatitis (NASH) via modulation of PPARα and SREBP-1. Food and chemical toxicology. 2013 Aug 31;58:198-209.
- 48) Sultana S, Perwaiz S, Iqbal M, Athar M. Crude extracts of hepatoprotective plants, Solanum nigrum and Cichorium intybus inhibit free radical-mediated DNA damage. Journal of ethnopharmacology. 1995 Mar 1;45(3):189-92.
- 49) Abd-EL-Rahim E, Yossef H, Soliman A. Natural additives for healthy crackers. Egyptian J. OF Nutrition. 2003;18:141-61.
- 50) Sabovics M, Gudreniece L, Kince T, Semjonovs P. Investigation of fructans increasing possibilities in rye bread. In11th Baltic Conference on Food Science and Technology" Food science and technology in a changing world" FOODBALT 2017, Jelgava, Latvia, 27-28 April 2017 2017 (pp. 96-101). Latvia University of Agriculture.
- 51) Zhu Z, Luo J, Ding L, Bals O, Jaffrin MY, Vorobiev E. Chicory juice clarification by membrane filtration using rotating disk module. Journal of food engineering. 2013 Mar 31;115(2):264-71.
- 52) Behera SS, Panda SH, Mohapatra S, Kumar A. Statistical optimization of elephant foot yam (Amorphophallus paeoniifolius) lacto-pickle for maximal yield of lactic acid. LWT-Food Science and Technology. 2018 Jan 1;87:342-50.